RESEARCH ARTICLE

Acute toxicity of *Galium* odoratum to the freshwater cladoceran *Moina* macrocopa

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Abstract

Galium odoratum (L.) is a medicinal plant with a number of health benefits, used in traditional and modern medicine. The toxicity of the coumarin in Galium odoratum is classified as high, however, no experimental data or data on toxic effects in humans following intake of Galium odoratum are available. The potential toxic effect can be estimated based on the content of coumarin and knowledge of its toxicity. The aim of the current study is to evaluate the acute toxicity effects of a range of concentrations of Galium odoratum water extract on Moina macrocopa and calculate the LC₅₀ within 24 hours. In order to compare the toxicity with those of other, well-known and widely used medicinal plants, extracts of Matricaria chamomilla and Tribulus terrestris are also tested.

The results show that LC_{50} value of *Galium odoratum* is comparable with those of *Matricaria chamomilla* and *Tribulus terrestris*, and *Galium odoratum* has intermediate toxicity between the two other studied species.

Keywords

Galium odoratum, Moina macrocopa, toxicity, LC₅₀

Introduction

Galium odoratum (L.) is a perennial herbaceous plant, naturally occurring in Europe, Asia and Northern Africa. It is used for centuries in traditional medicine for the treatment of central nervous system problems and disorders (Wszelaki et al., 2010; Kahkeshani et al., 2013; Friscic et al., 2018a), stomach, liver and digestive problems (Vlase et al., 2014; Mocan et al., 2016; Friscic et al., 2018a), gout treatment (Mocan et al.,

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2016), etc. It is valued for its antihypoxic (Iurchenko et al., 2015) and antimicrobial effect (Wagnera et al., 2017), ability to cleanse the blood (Friscic et al., 2018a), improve metabolism (Ilyina et al., 2016) and memory (Wszelaki et al., 2010), reduce swelling, cure wounds and cuts (Kahkeshani et al., 2013; Vlase et al., 2014; Wagnera et al., 2017).

Galium odoratum is used in modern medicine. Its proven pharmacological properties include antioxidant (Friscic et al., 2018b), antimicrobial, antifungal and antiviral effect (Cowan, 1999; Wojnicz et al., 2012; Vlase et al., 2014), burn wound healing activity (Kahkeshani et al., 2013), anti-hypoxic and sedative effect (Wojnicz et al., 2012; Iurchenko et al., 2015). The species is rich in biologically active compounds, including coumarin, asperulosiode, monotropein, scandoside, rutin, quercitrin, etc. (Wolf, 1993; Kahkeshani et al., 2013; Vlase et al., 2014).

The toxicity of the coumarin in *Galium odoratum* is classified as high (Duke, 1985; Cowan, 1999). The species is included in lists of toxic plants, e.g. Ordinance No. 1/12.01.2009, List of substances of FGFSA (2014), etc. However, no experimental data or data on toxic effects in humans following intake of *Galium* odoratum are available. The potential toxic effect can be estimated based on the content of coumarin and knowledge of its toxicity (BfR Health Assessment No 044, 2006; Egebjerg et al., 2018).

Cladoceran species are used in toxicity tests due to their short life cycle, easy cultivation in the laboratory, sensitivity to toxicants, small size requiring small volumes of extracts for testing, etc. Acute toxicity tests with *Moina macrocopa* are widely applied in ecotoxicology (Chu et al., 1997; Ji et al., 2008; Nam et al., 2009; Yi et al., 2010).

The aim of the current study is to evaluate the acute toxicity effects of a range of concentrations of *Galium odoratum* water extract on *Moina macrocopa* and calculate the LC_{50} (concentration lethal to 50% of the test animals) within 24 hours. In order to compare the toxicity with those of other, well-known and widely used medicinal plants, extracts of *Matricaria chamomilla* and *Tribulus terrestris* were also tested.

Materials and Methods

Galium odoratum aboveground biomass was collected in the summer of 2020 (during blossoming) from 8 localities in Mala and Ponor Mountains, Bulgaria. The row material was thoroughly mixed, air-dried and chopped. Dry aboveground biomass of *Matricaria chamomilla* and *Tribulus terrestris* was purchased from a recognized producer of Bulgarian medicinal plants.

50 g of dry plant material from each species were submerged in 1000 ml boiling water and left to macerate for 24 h. After maceration, the plant material was filtered, and the extracts were used for the preparation of the test solutions.

Series of concentrations were selected from the lists available in Report EPS 1/ RM/11 (1990). The tested concentrations are listed in Table 1.

For the toxicity test were used *Moina macrocopa* neonates (less than 24 hours old) from a laboratory culture maintained in a climate-controlled room at $25 \pm 2^{\circ}$ C, on a 16:8 h light/dark photoperiod for 2 weeks before testing. Animals were handled as

Extract (ml) in 100 ml test solution	Concentration (mg/ml) in the test solution		
1.00	0.50		
1.80	0.90		
3.20	1.60		
5.60	2.80		
10.00	5.00		
18.00	9.00		
32.00	16.00		
56.00	28.00		
100.00	50.00		

Table 1. Series of concentration used for the toxicity test

little as possible, carefully, and quickly to minimize stress. Dechlorinated municipal water was used for cultivation, dilution and control. During cultivation, animals were fed dry yeast once every 24 h.

Neonates for the tests were obtained as gravid females from the culture were transferred to another vessel 24 hours before the test and fed, in order to increase the number of reproducing animals. Neonates were held in the test solutions for 24 hours. They were not fed during the test. At the end of the test period, the living individuals in the control and test solutions were counted. The LC_{50} values and their 95% confidence intervals (CI) were calculated using Probit Analysis, following the methodology of Finney (1971) in Excel 2010 (Mekapogu, 2021). The data for mortality proportions were transformed to probits, and the respective concentrations – to log_{10} . The LC_{50} values were estimated using regression analysis (Busvine, 1971). The original mortality (observed) and derived mortality (expected) were used to calculate the Chi-Square test. The goodness of fit was assessed via the R² value.

Results and Discussion

The survival rate of *Moina macrocopa* in the control was 100%. The survival rates in the tested concentrations of *Galium odoratum*, *Matricaria chamomilla* and *Tribulus terrestris* are given in Table 2.

The mortality increased with the increase in the concentration of plant extracts according to the mortality concentration curves shown in Figure 1.

 LC_{50} values of the water extracts of the studied medicinal plants, calculated using probit analysis, are given in Table 3.

For all three plant species, the concentration of the extract had an inversely proportional relationship with the survival rate of *Moina macrocopa*. In the samples with concentrations 0.5 and 0.9 mg/ml, as well as in the control, the survival rate was 100%. In the *Matricaria chamomilla* extract, all water fleas survived also at 1.6 mg/ml and 2.8 mg/ml. In extracts with concentrations of 28 and 50 mg/ml there were no

Test solution	Survival rate (%)					
(mg/ml)	Galium odoratum	Matricaria chamomilla	Tribulus terrestris			
Control	100	100	100			
0.5	100	100	100			
0.9	100	100	100			
1.6	80	100	80			
2.8	80	100	60			
5	70	80	60			
9	60	60	0			
16	20	20	0			
28	0	0	0			
50	0	0	0			

Table 2. Survival rates for 24 h exposure of Moina macrocopa to aqueous extract of Galiumodoratum, Matricaria chamomilla and Tribulus terrestris

Table 3. LC 50 of Galium odoratum, Matricaria chamomilla, Tribulus terrestris

Smanias	LC ₅₀ (mg/ml)	95% CI		R ²
Species		Lower	Upper	R -
Galium odoratum	8.18	5.01	13.40	0.96
Matricaria chamomilla	11.59	7.83	17.16	0.97
Tribulus terrestris	6.74	3.71	12.20	0.93

living water fleas. In the extracts of *Tribulus terrestris* there were no living individuals also at concentrations 9 mg/ml and 16 mg/ml.

Based on the toxicity tests with *Moina macrocopa* can be concluded that *Galium odoratum* has intermediate toxicity between *Matricaria chamomilla* and *Tribulus terrestris*. The LC₅₀ of *Galium odoratum* is 8.18 mg/ml with a 95% CI 5.01-13.40 mg/ml. The highest toxicity of the three studied plant species was observed in *Tribulus terrestris* extracts. The LC₅₀ is 6.74, with a 95% confidence limit 3.71-12.20 mg/ml. *Matricaria chamomilla* showed the lowest toxicity with LC₅₀ 11.59 mg/ml with a 95% confidence limit of 7.83-17.16 mg/ml. Based on the results of the Chi-tests (non-significant), the observed and derived mortality showed a good fit.

Conclusion

The LC₅₀ value of *Galium odoratum* is 8.18 mg/ml with a 95% CI 5.01-13.40 mg/ml and is comparable with those of *Matricaria chamomilla* and *Tribulus terrestris*. It is 30% lower than the LC₅₀ of *Matricaria chamomilla* and 18% higher than this of *Tribulus terrestris*, i.e. the toxicity of *Galium odoratum* is higher than the toxicity of *Matricaria chamomilla* and lower than that of *Tribulus terrestris*.

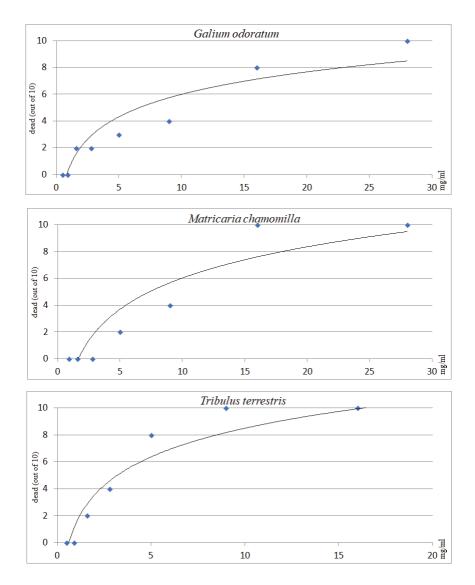


Figure 1. Mortality concentration curves (logarithmic trend line) for 24 h exposure of *Moina* macrocopa to aqueous extract of *Galium odoratum*, *Matricaria chamomilla* and *Tribulus terrestris*

References

- BfR Health Assessment No. 044. 2006. High daily intakes of cinnamon: Health risk cannot be ruled out.
- Boussadi, O., B. Martin, A. Nicolas, M. Bodson. 2015. Anthraquinone dyestuff and growth of *Galium odoratum* ((L.) Scop.) rhizomes in relation to ecophysiological and ontogenic conditions. – European Scientific Journal, 11(9), 1857-7881.
- Busvine, J.R. 1971. A critical review of the techniques for testing insecticides. Commonwealth Agricultural Bureaux, London, ISBN 0851980309.

- Chu, K.H., C.K. Wong, K.C. Chiu. 1997. Effects of the insects growth regulator (S)-Methoprene on survival and reproduction of the freshwater cladoceran *Moina macrocopa*.
 – Environmental Pollution, 96, 173-178.
- Cowan, M. 1999. Plant products as antimicrobial agents. Clinical Microbiology Reviews, 12(4), 564-582.
- Duke, J. A. 1985. Handbook of medicinal herbs, Boca Raton, CRC Press.
- Egebjerg, M., P. Olesen, F. Eriksen, G. Ravn-Haren, L. Bredsdorff, K. Pilegaard. 2018. Are wild and cultivated flowers served in restaurants or sold by local producers in Denmark safe for the consumer? – Food and Chemical Toxicity, 120, 129-142.
- Finney, D.J. 1971. Probit Analysis. 3rd Edition, Cambridge University Press, Cambridge.
- Friscic, M., M. Stibric Baglama, M. Milovic, K. Hazler Pilepic, Z. Males. 2018a. Traditional uses chemical composition and biological effects of *Galium L. species. – Farmaceutski* glasnik, 74(5), 343-350. (In Croatian).
- Friscic, M., M. Stibric Baglama, M. Milovic, K. Hazler Pilepic, Z. Males. 2018b. Content of bioactive constituents and antioxidant potential of *Galium* L. species. – Croatica Chemica Acta, 91(3), 1-7.
- Ilyina, T., A. Kovaleva, N. Yurchenko, N. Kashpur, A. Volyansky. 2011. Study of lipophilc compounds of the herb Asperula odorata L. and their biological activity. – Actual problems of pharmaceutical science and medical schools, from development to commercialization. Materials of a scientific/practical conference with international participation, dedicated to the 9th anniversary of the Perm State Pharmaceutical Academy, 85-87. (In Russian).
- Iurchenko, N., T. Ilyina, A. Kovaleva, E. Toryanik, I. Kulish .2015. Sedative and antihypoxic activity of Asperula odorata. – Pharmacognosy Communications, 5(4), 22-26.
- Ji, K., Y. Kim, S. Oh, B. Ahn, H. Jo, K. Choi. 2008. Toxicity of perfluorooctane sulfonic acid and perfluorooctanoic acid on freshwater macroinvertebrates (*Daphnia magna* and *Moina macrocopa*) and fish (*Oryzias latipes*). – Environmental Toxicology and Chemistry, 27, 2159–2168.
- Kahkeshani, N., B. Farahanikia, P. Mahdaviani, A. Abdolghaffari, Gh. Hasanzadeh, M. Abdollahi, M. Khanavi. 2013. Antioxidant and burn healing potential of *Galium odoratum* extracts. – Research in Pharmaceutical Sciences, 8, 197-203.
- List of substances of FGFSA. 2014. List of substances of the competent Federal Government and Federal State Authorities: category "Plants and Plant Parts". Springer.
- Mekapogu, A.R. 2021. Finney's probit analysis spreadsheet calculator (Version 2021). Available at: https://probitanalysis.wordpress.com/
- Mocan, A., G. Crişan, L. Vlase, B. Ivănescu, A. Sabin Bădărău, A. Letiția Arsene. 2016. Phytochemical investigations on four *Galium species* (Rubiaceae) from Romania. – Farmacia, 64(1), 95-99.
- Nam,, S.H., C.Y. Yang, Y.J. An. 2009. Effects of antimony on aquatic organisms (larva and embryo of Oryzias latipes, Moina macrocopa, Simocephalus mixtus, and Pseudokirchneriella subcapitata). – Chemosphere, 75, 889–893.
- Ordinance No. 1 of 12.01.2009 on the terms and conditions of the structure and safety of playgrounds. State gazette 10/2009, Ministry of Regional Development and Public Works, Bulgaria. (In Bulgarian).
- Report EPS 1/RM/11. 1990. Biological test method: Acute lethality test using Daphnia spp. (July 1990, amended May 1996). Method development and application section, Environmental Technology Centre, Canada. ISBN 0-662-18076-3.

- Vlase, L., A. Mocan, D. Hanganu, D. Benedec, A. Gheldiu, G. Crişan. 2014. Comparative study of polyphenolic content, antioxidant and antimicrobial activity of four *Galium species* (Rubiaceae). – Digest Journal of Nanomaterials and Biostructures, 9(3), 1085-1094.
- Wagnera, Ch., J. De Gezelle, M. Robertson, K. Robertson, M. Wilson, S. Komarnytsky. 2017. Antibacterial activity of medicinal plants from the physicians of Myddvai, a 14th century Welsh medical manuscript. – Journal of Ethnopharmacology, 203, 171-181.
- Wojnicz, D., A. Kucharska, A. Sokół-Łętowska, M. Kicia, D. Tichaczek-Goska. 2012. Medicinal plants extracts affect virulence factors expression and biofilm formation by the uropathogenic Escherichia coli. – Urological Research, 40(6), 683-697.
- Wolf, H. U. 1993. Hager's Textbook of Pharmaceutical Practice, Vol. 5, 5th ed. (Springer Heidelberg), 223.
- Wszelaki, N., A. Kuciun, A. Kiss. 2010. Screening of traditional European herbal medicines for acetylcholinesterase and butyrylcholinesterase inhibitory activity, Acta Pharmaceutica, 60, 119-128.
- Yi, X., S. Kang, J. Jung. 2010. Long-term evaluation of lethal and sublethal toxicity of industrial effluents using *Daphnia magna* and *Moina macrocopa*. – Journal of Hazardous Materials, 178, 1-3, 982-987.